



## **Identifying Possible Hepatic Fibrosis of Hepatitis B Origin Using Non-invasive Markers: A Case-control Study in the South West Region of Cameroon**

**Kukwah Anthony Tufon<sup>1,2\*</sup>, Henry Dilonga Meriki<sup>1,2,3</sup>,  
Kwenti Emmanuel Tebit<sup>1,2,3,4</sup>, Teuwafeu Denis Georges<sup>2,5</sup>,  
Nyeke James Tony<sup>1,2</sup>, Nicholas Tendongfor<sup>3</sup>, George Enow-Orock<sup>2,6</sup>  
and Damian Nota Anong<sup>7</sup>**

<sup>1</sup>*Department of Microbiology and Parasitology, University of Buea, Buea, South West Region, Cameroon.*

<sup>2</sup>*Buea Regional Hospital, Buea, Southwest Region, Cameroon.*

<sup>3</sup>*Department of Public Health and Hygiene, Faculty of Health Science, University of Buea, Buea, Southwest Region, Cameroon.*

<sup>4</sup>*Department of Medical Laboratory Science, Faculty of Health Science, University of Buea, Buea, Southwest Region, Cameroon.*

<sup>5</sup>*Department of Internal Medicine, Faculty of Health Science, University of Buea, Buea, Southwest Region, Cameroon.*

<sup>6</sup>*Department of Biomedical science, Faculty of Health Science, University of Buea, Buea, Southwest Region, Cameroon.*

<sup>7</sup>*Department of Biological science, Faculty of Science, University of Bamenda, Bamenda, North West Region, Cameroon.*

### **Authors' contributions**

*This work was carried out in collaboration between all authors. All authors read and approved the final manuscript.*

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## ABSTRACT

**Aim:** HBV infection is known to cause liver fibrosis as well as some extrahepatic manifestations. We aimed at assessing hematological changes and identifying possible hepatic fibrosis of Hepatitis B origin using non-invasive markers (NIMs).

**Study Design:** A hospital-based Case-control study

**Place and Duration of Study:** Conducted at the Buea Regional Hospital, South West Region of Cameroon from February 2016 to December 2017

**Methods:** We enrolled HBV infected treatment naïve patients and “healthy” controls. All participants were subjected to alanine aminotransferase (ALT) and aspartate aminotransferase (AST) measurement, Full blood count (FBC), HBsAg, anti-HBc, HIV and HCV tests. Aspartate-platelet ratio index (APRI), fibrosis based on 4 factors (FIB-4), age-platelet index (API) and AST/ALT ratio (AAR) were generated from the test results. A questionnaire was administered to collect demographic data, alcohol consumption and history of liver/kidney disease or metabolic syndrome.

**Results:** A total of 202 cases and 202 controls were enrolled. Hematocrit (HCT) was significantly higher ( $p < 0.001$ ) in cases than controls. The controls had significantly higher mean values for platelet ( $p = 0.005$ ), neutrophil ( $p = 0.032$ ) and number of individuals with AST/ALT ratio (AAR)  $\geq 1$ . Liver fibrosis was significantly associated with cases than controls based on APRI (OR:6.06, CI:3.59-10.24), FIB-4 (OR:5.35, CI:2.75-10.39) and API (OR:8.02, CI:1.81-35.55). Among the HBV infected cases, 69 (34.2%), 36(17.8%) and 8(4.0%) had results indicative of fibrosis from at least 2, at least 3 and all 4 NIMs respectively. AAR detected possible fibrosis in 136 HBV infected cases of which up to 77 (56.6%) were not detected as fibrosis by the other NIMs.

**Conclusion:** HBV infection affects neutrophil percentage, HCT, PLT, APRI, FIB-4 and API in our study population. AAR did not prove to be a reliable NIM. Using at least 3 NIMs for HBV infected patients can significantly scale up their reliability for determining liver fibrosis in clinical practice.

*Keywords: HBV infection; APRI; API; FIB-4; AST/ALT ratio; non-invasive markers; hematological changes; liver fibrosis.*

## ABBREVIATIONS

*AST-Aspartate Aminotransferase; ALT-Alanine Aminotransferase; AAR-AST/ALT Ratio; API-Age-Platelet Index; APRI-Aspartate-Platelet Ratio Index; FBC-Full Blood Count; FIB-4-Fibrosis Index Based on 4 Factors; Anti-HBc-Hepatitis B Core Antibody; HBsAg-Hepatitis B Surface Antigen; HBV-Hepatitis B Virus; HCC- Hepatocellular Carcinoma; HCT-Hematocrit; HCV-Hepatitis C Virus; HIV-Human Immunodeficiency Virus; NIM-Non-Invasive Markers; PLT-Platelets; RBC-Red Blood Cells; SPSS-Statistical Package for Social Sciences; SW-South West; ULN-Upper Limit Normal; WBC-White Blood Cells; OR-Odds ratio; CI-Confidence Interval.*

## 1. INTRODUCTION

Hepatitis B virus (HBV) infection is known to be a major cause of liver cirrhosis worldwide. The risk of liver cancer is greatly increased once cirrhosis develops. Cirrhosis and liver cancer are now among the top ten causes of death worldwide [1,2]. Monitoring of chronic liver disease to identify and assess the stage of fibrosis is of paramount importance in preventing cirrhosis.

Liver biopsy is known to be the gold standard method for assessing liver fibrosis [3] although it is costly, invasive, has increased risk of complication, requires hospitalization, could lead to underestimation of fibrosis stage and has inter- and intra-observer discrepancies [4–8]. The

rapid development of new medications for the treatment of liver diseases (including chronic hepatitis B) increases the requirement for more frequent evaluation of liver fibrosis to assess treatment response. Liver biopsies are not ideal for frequent evaluations and thus cannot be regularly used for this purpose. HBV infected patients who require treatment usually have a higher Aspartate-Platelet Ratio Index (APRI) score as compared to those who don't require treatment [9]. This justifies the need to as well rely on non-invasive markers (NIMs) in evaluating hepatic fibrosis and determining treatment necessity [10].

NIMs for hepatic fibrosis must be simple, readily available, reliable, inexpensive, safe, and well

validated in different forms of chronic liver disease [11]. Some NIMs include Aspartate-platelet ratio index (APRI) score [12,13], Age-platelet index (API) [14], Fibrosis index based on 4 factors (FIB-4) [15,16] and AST/ALT ratio (AAR) [17,18].

The use of APRI and FIB-4 as non-invasive markers (NIMs) have been validated [12] but still not very much used in Cameroon despite the high prevalence of HBV infection. Liver biopsy is not commonly done in most parts of the country due to limited number of specialists who can carry out the procedure. As a result, liver fibrosis assessment in Cameroon happens to be a significant loop hole in the management of HBV infected patients.

Most of the studies that have been done so far to investigate the use of non-invasive markers actually enrolled just HBV infected patients with a recent liver biopsy result which was used as the gold standard to compare with and ascertain the use of the non-invasive markers. These studies did not address the possibilities of excluding or including fibrosis in HBsAg negative people using NIMs. In order to investigate how well these non-invasive markers could be used in excluding fibrosis or liver damage especially of hepatitis B origin, we compared the scores of some non-invasive markers (APRI, API, FIB-4 and AAR) between HBV infected cases and “healthy” controls.

The complex nature of HBV infection and the fact that it affects the liver gives the infection the potentials to indirectly account for extrahepatic manifestations causing blood related conditions like anemia [19–21] and significant changes in white blood cells (WBC) [22]. This study also investigated the effects of HBV infection on some hematological parameters so as to identify hematological changes that may occur in HBV chronically infected patients in the South West Region of Cameroon.

## 2. MATERIALS AND METHODS

### 2.1 Study Design and Sampling Technique

This was a hospital-based case-control study that enrolled HBV infected individuals and “healthy” controls all  $\geq 18$  years of age. The study was conducted in Buea, the capital city of the South West region of Cameroon. The cases were HBV treatment naïve hepatitis B surface antigen (HBsAg) positive people who were

negative for HIV and HCV with no history of kidney disease or metabolic syndrome. The controls were physically strong and “healthy” looking people negative for HIV, HCV, HBsAg and hepatitis B core antibody (anti-HBc) with no history of liver disease, kidney disease or metabolic syndrome. Alcohol consumers as well as pregnant women were excluded from the study. The participants were enrolled during an HIV, HCV and HBV free screening exercise we conducted in the Buea Regional Hospital. People who came to donate blood and tested positive for HBsAg were as well enrolled as cases.

### 2.2 Sample Size Calculation

Case-control study (80% power desired with an Odds ratio of  $\geq 2.0$ ). We used the formula in equation 1 below:

$$n = \left(\frac{r+1}{r}\right) \frac{(p)(1-p)(Z_{\beta}+Z_{\alpha/2})^2}{(P_1-P_2)^2} \quad [23] \quad (1)$$

Where

n=minimum sample size to achieve 80% power

r=ratio of controls to cases (1)

$Z_{\beta}$ = the desired power (typically .84 for 80% power).

$Z_{\alpha/2}$ =the desired level of statistical significance (typically 1.96).

$p(1-p)$  = A measure of variability (similar to standard deviation)

$P_1$ =Proportion of controls with elevated ALT: 21.7% [24]

$P_2$ =Proportion of HBV infected cases with elevated ALT

$$P_2 = \frac{ORP_{control\ exp}}{P_{control\ exp}(OR-1)+1} = \frac{2.0(0.22)}{0.22(2.0-1)+1} = 0.36$$

Therefore

$$n = 2 \frac{0.29(1-0.29)(0.84+1.96)^2}{(0.36-0.22)^2} = 164.7$$

Therefore, at least 165 controls and 165 HBV infected cases were considered to achieve the desired power (80%).

### 2.3 Data Collection and Clinical Evaluation

A standard questionnaire [25] was administered (interview mode) to all participants to obtain demographic information as well as to find out if

the participants consume alcohol and if they have any history of liver disease, kidney disease or metabolic syndrome. A medical doctor clinically examined the participants for signs of metabolic syndrome and/or kidney disease.

## 2.4 Sample Collection

Using the vacutainer system, a phlebotomist collected 10 ml of blood from each participant into 2 tubes: one containing Potassium Ethylenediaminetetraacetic acid (K<sub>3</sub>EDTA) anticoagulant and the other with no anticoagulant (dry tube).

## 2.5 Laboratory Analysis

### 2.5.1 Full Blood Count (FBC) and WBC Differential Count

The sample in the K<sub>3</sub>EDTA tube was used to perform full blood count (FBC) using an Auto Hematology Analyzer (Mindray model BC-2800, Mindray Bio-Medical Electronics, Nanshan, Shenzhen, P.R. China) following manufacturer's instructions. We considered the reference ranges as stated in the Mindray model BC-2800 operator's manual. WBC differential count was done manually for each blood sample in K<sub>3</sub>EDTA tube following a standard procedure [26]. We considered reference ranges as 20-40% and 40-80% for lymphocyte and neutrophil respectively [26].

### 2.5.2 Immunochromatographic qualitative tests

The samples in the dry tubes were centrifuged at 1000g for 5 minutes to obtain sera which was first of all used to screen for HIV (Abbot Determine, USA), HCV antibodies (Acon@ Laboratories Inc., USA) and HBsAg (Diaspot Inc. USA). HBV serologic profile was done using a qualitative panel kit (Blue Cross Bio-Medical Co. Beijing) for the detection of HBsAg, anti-HBs, HBeAg, anti-HBe and anti-HBc following manufacturer's instructions

### 2.5.3 Liver Aminotransferase measurement

With the sera, liver aminotransferase was measured for each participant using AST-(GOT)-Human reagent and ALT-(GPT)-Human reagent by spectrophotometry (Mindray® BA-88 Biochemistry analyser) following the manufacturer's instructions. ALT reference ranges were <32 IU/ml for females and <42 IU/ml for males while AST reference ranges were <31 IU/ml for females and <37 IU/ml for males as per the reagent used.

## 2.6 Calculation and Interpretation of Non-invasive Markers (NIMs)

### 2.6.1 APRI score

This was calculated by inserting the corresponding AST level, Upper limit normal (ULN) AST value and Platelet count in the following formula (equation 2):

$$APRI = \frac{AST\ level/ULN*}{Platelet\ count(10^9/L)} \times 100 \quad [27] \quad (2)$$

\*ULN is the upper limit normal of AST for the reagent used.

An APRI score <0.5 and ≥0.5 was used to exclude and include possible fibrosis respectively [12].

### 2.6.2 FIB-4 score

FIB-4 was calculated by inserting age, AST value, ALT value and platelet count in the following formula (equation 3):

$$FIB\ 4 = \frac{Age \times AST}{Platelet\ count \times \sqrt{ALT}} \quad [28] \quad (3)$$

A FIB-4 value of <1.45 and ≥1.45 was used to exclude and include possible fibrosis respectively [28,29]

### 2.6.3 API score

Age-platelet index was obtained using age grouped as follows: <30 years old; 0 points, 30-39 years old; 1 point, 40-49 years old; 2 points, 50-59 years old; 3 points, 60-69 years old; 4 points and ≥70 years old: 5 points. The platelet count (10<sup>3</sup>/μl) was grouped as follows: ≥225; 0 points, 200-224; 1 point, 175-199; 2 points, 150-174; 3 points, 125-149; 4 points and <125; 5 points [14]. It was then calculated using the following formula (equation 4):

$$API = (points\ for\ age) + (Points\ for\ Platelet) \quad [14] \quad (4)$$

An API score ≥6 indicated significant histologic findings (necroinflammatory lesions, septal fibrosis and/or cirrhosis). Scores <6 showed no fibrosis [14,30].

### 2.6.4 Aspartate Aminotransferase/Alanine Aminotransferase Ratio (AAR)

This was calculated using the formula (equation 5):

$$AAR = \frac{AST}{ALT} \quad [31] \quad (5)$$

AAR < 1 and AAR ≥ 1 was used to exclude and include significant fibrosis respectively [30].

## 2.7 Statistical Analysis

Data was entered in MS excel and analyzed in SPSS (Statistical Package for Social Sciences) version 21. Data were presented using frequency distribution tables and summary statistics. Categorical variables were compared using the Pearson's Chi-square test or the Fisher's exact test. Continuous variables were compared using the student t-test. A logistic regression analysis

was fitted for cases that recorded p value < 0.05 in the crude Odds ratio. A p value < 0.05 was considered significant for all analyses.

## 3. RESULTS

After matching for age and sex, a total of 404 participants (202 HBsAg positive individuals and 202 "healthy" controls) were enrolled. Mean ages of 31.39±8.3 and 31.89±8.5 were recorded for the cases and controls respectively. Cases had 100(49.5%) females and 102 (50.5%) males while controls had 104 (51.5%) females and 98(48.5) males. All the participants were aged between 18 and 65 years (Table 1).

**Table 1. Age and gender distribution**

Groups (n)	Mean age (years)	Age range (years)	Gender	n (%)
HBV infected cases (202)	31.39±8.3	18-62	Female	100 (49.5)
			Male	102 (50.5)
Healthy controls (202)	31.89±8.5	18-65	Female	104 (51.5)
			Male	98 (48.5)

**Table 2. Mean comparisons of liver enzymes, hematological parameters and non-invasive markers (NIMs) between HBV infected cases and controls**

Test group	Parameter	Status	Mean±SD	95% CI*	P-value
Liver enzymes	ALT (U/L)	Cases	32.6±18.8	8.3 to 14.0	<0.001
		Controls	21.5±7.7		
	AST (U/L)	Cases	35.5±23.4	7.2 to 14.0	<0.001
		Controls	24.9±7.8		
Hematological parameters	HGB (g/dl)	Cases	14.2±2.1	1.08 to 1.48	0.759
		Controls	14.4±9.0		
	WBC (10 <sup>3</sup> /μl)	Cases	5.0±1.4	-0.25 to 0.25	1.000
		Controls	5.0±1.1		
	RBC (10 <sup>6</sup> /μl)	Cases	5.0±0.6	0.02 to 0.22	0.095
		Controls	4.9±0.6		
	Lymphocyte (%)	Cases	44.7±10.7	0.76 to 2.96	0.247
		Controls	43.6±8.2		
	Neutrophil (%)	Cases	50.0±11.8	0.20 to 4.41	0.032
		Controls	52.3±9.6		
HCT (%)	Cases	42.9±6.2	1.01 to 3.19	<0.001	
	Controls	40.8±4.9			
PLT (10 <sup>3</sup> /μl)	Cases	220.3±62.4	5.08 to 28.92	0.005	
	Controls	237.3±59.4			
Non-invasive markers of fibrosis	APRI	Cases	0.50±0.32	0.12 to 0.22	<0.001
		Controls	0.33±0.12		
	FIB-4	Cases	0.97±0.6	0.09 to 0.29	<0.001
		Controls	0.78±0.4		
	AAR	Cases	1.18±0.4	-0.04 to 0.10	0.394
		Controls	1.21±0.3		
	API	Cases	2.06±1.8	0.03 to 0.67	0.030
		Controls	1.71±1.4		

\*confidence interval

**Table 3. Comparing the interpretation of liver enzymes and hematological parameters between HBV infected cases and controls**

Test group	Parameter	Status	Interpretation			P-value
			Low	Normal	High	
Liver enzymes	ALT (U/L)	Cases	-	150 (74.2)	52 (25.8)	<b>&lt;0.001</b>
		Controls	-	189 (93.6)	13 (6.4)	
	AST (U/L)	Cases	-	135 (66.8)	67 (33.2)	<b>&lt;0.001</b>
		Controls	-	183 (90.6)	19 (9.4)	
Hematological parameters	HGB (g/dl)	Cases	25 (12.4)	177 (87.6)	-	0.426
		Controls	34 (16.8)	168 (83.2)	-	
	WBC ( $10^3/\mu\text{l}$ )	Cases	31 (15.3)	171 (84.7)	-	0.421
		Controls	37 (18.3)	165 (81.7)	-	
	RBC ( $10^6/\mu\text{l}$ )	Cases	4 (2.0)	139 (68.8)	59 (29.2)	0.773
		Controls	3 (1.5)	144 (71.3)	55 (27.2)	
	Lymphocyte (%)	Cases	2 (1.0)	73 (36.1)	127 (62.9)	0.398
		Controls	2 (1.0)	82 (40.6)	118 (58.4)	
	Neutrophil (%)	Cases	102 (50.5)	95 (47.0)	5 (2.5)	<b>0.041</b>
		Controls	67 (33.2)	133 (65.8)	2 (1.0)	
	HCT (%)	Cases	33 (16.3)	161 (79.7)	8 (4.0)	0.049
		Controls	44 (21.8)	157 (77.7)	1 (0.5)	
PLT ( $10^3/\mu\text{l}$ )	Cases	27 (13.4)	174 (86.1)	1 (0.5)	<b>&lt;0.001</b>	
	Controls	3 (1.5)	193 (95.5)	6 (3.0)		

**Table 4. Comparing the interpretation of non-invasive markers for fibrosis between HBV infected cases and controls**

Non-invasive markers	Status	NIM interpretation		Crude odds ratio (CI)	P-value	Adjusted odds ratio (CI)	P-value
		Possible fibrosis	No fibrosis				
APRI	Cases, n (%)	86 (42.6)	116 (57.4)	6.06 (3.59-10.24)	<0.001	1.77 (1.47-2.13)	<0.001
	Controls, n (%)	22 (10.9)	180 (89.1)				
FIB-4	Cases, n (%)	51 (25.2)	151 (74.8)	5.35 (2.75-10.39)	<0.001	6.06 (2.92-12.50)	<0.001
	Controls, n (%)	12 (5.9)	190 (94.1)				
API	Cases, n (%)	15 (7.4)	187 (92.6)	8.02 (1.81-35.55)	0.006	7.58 (1.54-37.04)	0.013
	Controls, n (%)	2 (1.0)	200 (99.0)				
AAR	Cases, n (%)	136 (67.3)	66 (32.7)	0.48 (0.30-0.76)	0.002	0.52(0.32-0.83)	0.006
	Controls, n (%)	164 (81.2)	38 (18.8)				

(APRI <0.5=No fibrosis, APRI ≥0.5=Possible fibrosis: FIB-4 <1.45=No fibrosis, FIB-4≥1.45 =Possible fibrosis: API<6= No fibrosis, API ≥6=Possible fibrosis: AAR<1=No fibrosis, AAR≥1=Possible fibrosis)

A statistically significant difference (P<0.05) was recorded for the mean values of ALT, AST, neutrophil, HCT, PLT, APRI, FIB-4 and API between cases and controls (Table 2). Cases recorded a significantly higher number of participants with abnormal results for ALT, AST, neutrophil and platelet as seen in Table 3. The

mean AAR (p=0.394) did not show any statistically significant difference between cases and controls (Table 2) although there were significantly more controls (p=0.002) with AAR≥1 (Table 4). Liver fibrosis was significantly more associated with cases than controls based on APRI (OR:6.06, CI:3.59-10.24), FIB-4 (OR:5.35,

CI:2.75-10.39) and API (OR:8.02, CI:1.81-35.55) both in the crude and adjusted Odds ratio (Table 4).

Considering APRI  $\geq 0.5$  and FIB-4  $\geq 1.45$  as the low cut-offs to denote fibrosis [12,28] for our HBV infected participants, 86 (42.6% of 202) and 51 (25.2% of 202) of them were considered to have some form of fibrosis at least based on APRI and FIB-4 respectively. AAR recorded 136 (67.3% of 202) cases with possible fibrosis while API had 15 (7.4% of 202). The 4 NIMs excluded fibrosis in 27 (13.4%) of the 202 HBV infected cases as seen in Fig. 1. Seventy-seven (38.1%) and 29 (14.4%) of the cases showed evidence of fibrosis based on AAR only and APRI only respectively.

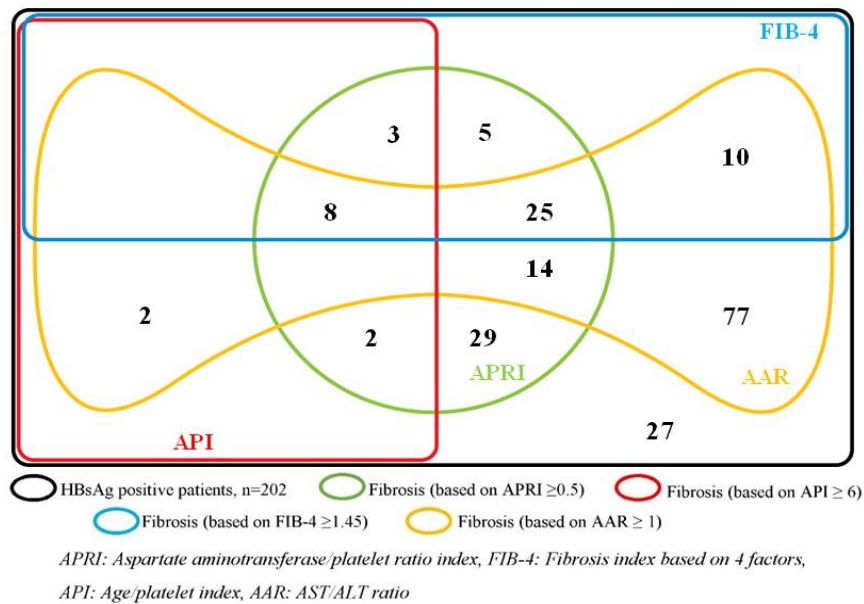
Sixty-nine (34.2%) of the cases had a result indicative of possible fibrosis from at least 2 of the different NIMs (Fig. 1). Thirty-six (17.8%) had an interpretation indicative of possible fibrosis from at least 3 of the different NIMs while 8(4.0%) had an interpretation indicative of fibrosis from all the 4 different NIMs (Fig. 1).

#### 4. DISCUSSION

HBV is known to affect mainly the liver but the inflammatory and immune response in HBV infected patients can affect the clinical, serological and hematological outcome of the patient as the severity of the viral disease depends on the immune system's ability to attack

infected hepatocytes [32]. Our study looked at possible changes in some hematological parameters (HB, RBC, WBC, PLT, HCT, Neutrophil and lymphocyte) that may have been brought about by HBV infection. Cases recorded a significantly lower mean neutrophil percentage as compared to controls. Some other studies showed a statistically significant difference in the mean Hemoglobin measurements of cases and controls [33] while others did not have a statistical difference for neutrophil but rather had a statistical difference for lymphocytes [22] with the mean lymphocyte percentage of HBV infected cases significantly higher than that of the controls. This may look different from our findings but the clinical implication and interpretation is similar because during viral infections, a rise in lymphocyte percentage and a drop in neutrophil percentage is usually noticed [34].

Our study showed that the mean platelet count in HBV infected cases was significantly lower than that of the controls as seen in other studies [35,36]. Thrombocytopenia (deficiency of platelets in the blood) is a common complication in liver disease. The major mechanisms for thrombocytopenia in liver cirrhosis are (1) platelet sequestration in the spleen; and (2) decreased production of thrombopoietin in the liver. More platelets are destroyed in the enlarging spleen as a result of fibrosis and aggravated portal hypertension [37].



**Fig. 1. Relationship and overlap between different non-invasive markers of liver fibrosis in the HBV infected cases (n=202)**

HBV infected cases recorded a significantly higher HCT as compared to controls in our study. This does not corroborate with previous findings [22,33,38]. We did not notice any statistically significant difference with the mean values of hemoglobin and RBC between cases and controls. Although some studies [19,20] have shown that HBV infection can cause anemia, the infection is rarely linked to hematological changes in erythrocyte or its indices. Most studies link it to changes in leucocytes and platelets in blood [38].

Over the past decade several non-invasive markers of fibrosis have been proposed and validated following research. A good number of them are quite inexpensive and does not require a lot of skills like the liver biopsy which is presently still considered as the gold standard despite all its short comings. The mean APRI, FIB-4 and API scores of HBV infected cases were significantly higher than that of the controls in our study. Similar findings were reported in previous studies with APRI [39].

AAR (AST/ALT ratio) has been widely utilized as a predictor of hepatic fibrosis [40]. We did not notice any statistically significant difference between the mean AAR of HBV infected cases and controls (Table 4). Interestingly there was significantly more “healthy” controls with  $AAR \geq 1$  (indicative of possible fibrosis) than HBV infected cases (Table 4). AST is seemingly more abundant in cells (as compared to ALT) because it can be found in the heart, liver, kidney, brain, RBC and muscle tissues whereas ALT is mainly found in liver cells [41]. Moreover, even at the level of the liver, ALT is only found in the cytoplasm of hepatocytes whereas AST can be found in the cytoplasm as well as in the mitochondria of hepatocytes [42]. Elevation of AST above ALT (which would probably result to  $AAR \geq 1$ ) may be seen in several different conditions. It could be suggestive of a non-hepatic source of AST which may occur due to release of AST from blood cells during hemolysis or reduction in AST clearance [43]. AST may as well be released into the circulation following injury to non-liver cells in the body (muscle, heart, kidney, brain etc) particularly cells that contain mitochondria [42]. This lack of specificity probably explains why our study recorded a significant proportion of HBsAg negative people with  $AAR \geq 1$ .

Some researchers now criticize and contest the use of AAR (AST/ALT ratio) as a marker of

fibrosis especially in chronic HBV infection [44,45]. AST/ALT ratios below 1.0 ( $AAR < 1$ ) are typical of chronic viral hepatitis (e.g. hepatitis B and C). However ratios slightly above 1.0 may be found in chronic viral hepatitis but this is particularly when progression to fibrosis and cirrhosis is present [42]. Our Study couldn't agree less as we recorded a higher number of HBV infected cases with  $AAR < 1$  [66 (32.7%)] as compared to controls with  $AAR < 1$  [38 (18.8%)]. Some studies have shown that AAR is a better marker for fibrosis in Alcoholic hepatitis and not viral hepatitis [41,46].

Based on the fact that cut off values for many non-invasive markers are still under debate, a high cut off with high specificity (i.e. fewer false-positive results) and a low cut off with high sensitivity (i.e. fewer false-negative results) have been generated for APRI and FIB-4 [12,47] to include and exclude fibrosis respectively. This controversy on cut off values actually questions the reliability of NIMs in determining liver fibrosis. In order to bypass this problem, we tried to compare the relationship and/or overlap between 4 different NIMs under the assumption that if all the NIMs or most of the NIMs share the same conclusion for a patient, then its most likely the true clinical picture of the patient as far as possible presence or absence of liver fibrosis is concerned. We realized that there was no individual who had a FIB-4 or API score indicative of fibrosis based on FIB-4 or API only unlike APRI (independently showed possible fibrosis in 29 cases) and AAR (independently showed possible fibrosis in 77 cases). The fact that APRI alone identified possible fibrosis in 29 cases which were not identified as such by the other NIMs further confirms previous findings which showed that the  $< 0.5$  cut off mark for APRI was not very reliable in ruling out fibrosis [47–49] as compared to the  $< 1.45$  cut off for FIB-4. In our study, 34.2%, 17.8% and 4.0% of our HBV infected cases had results indicative of possible hepatic fibrosis with at least 2 NIMs, at least 3 NIMs and all 4 NIMs respectively. A systematic review that assessed the proportion of HBV patients with liver fibrosis in Sub Saharan Africa showed that over 15% have significant fibrosis/cirrhosis [50]. A study conducted in Cameroon recorded 13.4% of HBV patients with fibrosis [51]. Another study conducted in the Gambia had 17.5% prevalence of fibrosis among HBV infected individuals screened at a blood bank. These proportions of HBV infected patients with liver fibrosis are similar to the 17.8% of patients we had with fibrosis based on



at least 3 different NIMs. Although this may not be a solid foundation to validate the use of NIMs, it could ascertain their reliability in our setting if at least 3 of them are considered before ruling in or ruling out fibrosis for every HBV infected patient on clinical assessment. Unfortunately, liver biopsy was not done in this study which could have been used to further justify the use of NIMs in our setting.

## 5. CONCLUSION

HBV infection affects neutrophil, HCT, PLT, APRI, FIB-4 and API in our study population. AAR did not prove to be a reliable NIM in detecting fibrosis of hepatitis B origin due to lack of specificity. Using more than two NIMs for an HBV infected patient at a given time can by-pass the problem of contradictory cut off values and significantly reduce the possibilities of arriving at a wrong conclusion when determining fibrosis in clinical practice. Moreover, most NIMs are inexpensive and may not pose that much of a problem financially if the doctor demands for more than one at a given time. Besides some of the NIMs share similar parameters and as such, data generated for one may as well be used for another like APRI and FIB-4.

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## CONSENT

A signed informed consent form was obtained from every participant prior to enrolment. A trained counsellor verbally explained the study protocol to those who could not read nor write and counselled each participant prior to testing.

## ETHICAL APPROVAL

The National Ethics Committee of Research for Human Health (NECRHH) in Cameroon approved the study protocol.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Griffiths C, Rooney C, Brock A. Leading causes of death in England and Wales—how should we group causes? *Health Stat Q.* 2005;6–17.
2. Bosetti C, Levi F, Lucchini F, Zatonski W, Negri E, La Vecchia C. Worldwide mortality from cirrhosis: An update to 2002. *J Hepatol.* 2007;46:827–839.
3. Poynard T, Imbert-Bismut F, Munteanu M, Messous D, Myers RP, Thabut D, et al. Overview of the diagnostic value of biochemical markers of liver fibrosis (FibroTest, HCV FibroSure) and necrosis (ActiTest) in patients with chronic hepatitis C. *Comp Hepatol.* BioMed Central. 2004;3: 8.  
DOI: 10.1186/1476-5926-3-8
4. Cadranet J, Rufat P, Degos F. Practices of liver biopsy in France: Results of a prospective nationwide survey. For the Group of Epidemiology of the French Association for the Study of the Liver (AFEF). *Hepatology.* 2000;32:477–481.
5. Wong JB, Koff RS. Watchful waiting with periodic liver biopsy versus immediate empirical therapy for histologically mild chronic hepatitis C. A cost-effectiveness analysis. *Ann Intern Med.* 2000;133:665–675.  
Available:  
<http://www.ncbi.nlm.nih.gov/pubmed/11074899>
6. Afdhal NH, Nunes D. Evaluation of liver fibrosis: A concise review. *Am J Gastroenterol.* 2004;99:1160–1174.  
DOI: 10.1111/j.1572-0241.2004.30110.x
7. Westin J, Lagging LM, Wejstål R, Norkrans G, Dhillon AP. Interobserver study of liver histopathology using the Ishak score in patients with chronic hepatitis C virus infection. *Liver.* 1999;19:183–187.  
Available:<http://www.ncbi.nlm.nih.gov/pubmed/10395036>
8. Regev A, Berho M, Jeffers LJ, Milikowski C, Molina EG, Pyrsopoulos NT, et al. Sampling error and intraobserver variation in liver biopsy in patients with chronic HCV infection. *Am J Gastroenterol.* 2002;97: 2614–2618.  
DOI: 10.1111/j.1572-0241.2002.06038.x
9. Tufon KA, Anong DN, Meriki HD, Georges TD, Maurice M, Kouanou YS, et al. Characterization and assessment of HBV chronically infected patients: Identification of those eligible for treatment in the South

- West region of Cameroon. PLoS One. Public Library of Science. 2018;13: e0203312.  
DOI: 10.1371/journal.pone.0203312
10. Castera L, Pinzani M. Biopsy and non-invasive methods for the diagnosis of liver fibrosis: Does it take two to tango? Gut. 2010;59: 861–866.  
DOI: 10.1136/gut.2010.214650
  11. Lee S, Kim DY. Non-invasive diagnosis of hepatitis B virus-related cirrhosis. World J Gastroenterol. 2014;20:445.  
DOI: 10.3748/wjg.v20.i2.445
  12. WHO. Guidelines for the prevention, care and treatment of persons with chronic hepatitis b infection [Internet]; 2015. Available: [http://apps.who.int/iris/bitstream/10665/154590/1/9789241549059\\_eng.pdf?ua=1&ua=1](http://apps.who.int/iris/bitstream/10665/154590/1/9789241549059_eng.pdf?ua=1&ua=1)
  13. Terrault NA, Bzowej NH, Chang KM, Hwang JP, Jonas MM, Murad MH. AASLD guidelines for treatment of chronic hepatitis B. Hepatology. 2016;63:261–283.  
DOI: 10.1002/hep.28156
  14. Poynard T, Bedossa P. Age and platelet count: A simple index for predicting the presence of histological lesions in patients with antibodies to hepatitis C virus. J Viral Hepat. Blackwell Publishing Ltd. 1997;4: 199–208.  
DOI: 10.1046/j.1365-2893.1997.00141.x
  15. Sterling RK, Lissen E, Clumeck N, Sola R, Correa MC, Montaner J, et al. Development of a simple noninvasive index to predict significant fibrosis in patients with HIV/HCV coinfection. Hepatology. 2006;43:1317–1325.  
DOI: 10.1002/hep.21178
  16. Li Y, Chen Y, Zhao Y. The diagnostic value of the FIB-4 index for staging hepatitis B-related fibrosis: A meta-analysis. PLoS One. 2014;9.  
DOI: 10.1371/journal.pone.0105728
  17. McPherson S, Stewart SF, Henderson E, Burt AD, Day CP. Simple non-invasive fibrosis scoring systems can reliably exclude advanced fibrosis in patients with non-alcoholic fatty liver disease. Gut. 2010; 59:1265–1269.  
DOI: 10.1136/gut.2010.216077
  18. Fallatah HI, Fallatah HI, Fallatah IH. Noninvasive biomarkers of liver fibrosis: An overview. Adv Hepatol. 2014;2014:1–15.  
DOI: 10.1155/2014/357287
  19. Bozkaya H, Yurdaydin C, Törüner M, Arat M, Bozdayi AM, Ereku S, et al. Remission of severe aplastic anemia associated with hepatitis B virus infection after viral clearance: potential role of lamivudine. Dig Dis Sci. 2002;47:1782–1785. Available: <http://www.ncbi.nlm.nih.gov/pubmed/12184530>
  20. Qureshi K, Sarwar U, Khallafi H, Qureshi K, Sarwar U, Khallafi H. Severe aplastic anemia following acute hepatitis from toxic liver injury: Literature review and case report of a successful outcome. Case Reports Hepatol. 2014;1–7.  
DOI: 10.1155/2014/216570
  21. Adinolfi LE, Giordano MG, Andreana A, Tripodi MF, Utili R, Cesaro G, et al. Hepatic fibrosis plays a central role in the pathogenesis of thrombocytopenia in patients with chronic viral hepatitis. Br J Haematol. 2001;113:590–595. Available: <http://www.ncbi.nlm.nih.gov/pubmed/11380442>
  22. Eze M, Buseri F, WAchukwu C, Nnatuanya I. Effects of hepatitis B infection on haematological parameters in pregnancy in Port Harcourt, Nigeria. Res J Med Sci. 2009;3:194–197.
  23. Charan J, Biswas T. How to calculate sample size for different study designs in medical research? Indian J Psychol Med. Wolters Kluwer- Medknow Publications. 2013;35:121–6.  
DOI: 10.4103/0253-7176.116232
  24. Abongwa LE, Akono N, Elo AP, Vershiyi C. Incidence of elevated aminotransferases levels among patients presenting with hepatitis syndrome in the northwest region of Cameroon. Cohesive J Microbiol Infect Dis. 2018;1:5–9.  
DOI: 10.31031/CJMI.2018.01.000504
  25. Meriki HD, Tufon KA, Anong DN, Tony NJ, Kwenti TE, Bolimo AF, et al. Vaccine uptake and immune responses to HBV infection amongst vaccinated and non-vaccinated healthcare workers, household and sexual contacts to chronically infected HBV individuals in the South West Region of Cameroon. Bansal GP, editor. PLoS One. Public Library of Science. 2018;13: e0200157.  
DOI: 10.1371/journal.pone.0200157
  26. CLSI. H20-A2 Reference Leukocyte (WBC) differential count (proportional) and evaluation of instrumental methods; 1992.
  27. Wai CT, Greenson JK, Fontana RJ, Kalbfleisch JD, Marrero JA, Conjeevaram HS, et al. A simple noninvasive index can predict both significant fibrosis and

- cirrhosis in patients with chronic hepatitis C. *Hepatology*. 2003;38: 518–526.  
DOI: 10.1053/jhep.2003.50346
28. Li Q, Ren X, Lu C, Li W, Huang Y, Chen L. Evaluation of APRI and FIB-4 for noninvasive assessment of significant fibrosis and cirrhosis in HBeAg-negative CHB patients with ALT  $\geq$  2 ULN. 2017;12:1–7.  
DOI: 10.1097/MD.0000000000006336
  29. Kim BK, Kim DY, Park JY, Ahn SH, Chon CY, Kim JK, et al. Validation of FIB-4 and comparison with other simple noninvasive indices for predicting liver fibrosis and cirrhosis in hepatitis B virus-infected patients. *Liver Int*. 2010;30:546–553.  
DOI: 10.1111/j.1478-3231.2009.02192.x
  30. Atayan Y. The comparison of liver fibrosis score and non-invasive tests in naive chronic viral hepatitis B patients. 2017;28:7790–7792.
  31. Giannini E, Risso D, Botta F, Chiarbonello B, Fasoli A, Malfatti F, et al. Validity and clinical utility of the aspartate aminotransferase-alanine aminotransferase ratio in assessing disease severity and prognosis in patients with hepatitis C virus-related chronic liver disease. *Arch Intern Med*. 2003;163:218–24.  
Available: <http://www.ncbi.nlm.nih.gov/pubmed/12546613>
  32. Ajugwo AO, Ukaji DC, Erhabor TA, Adias TC. Some haematological parameters of symptomatic and asymptomatic hepatitis B positive patients attending a Nigerian Tertiary Hospital. 2015;7:219–223.  
DOI: 10.9734/BJMMR/2015/15491
  33. Uche OJCI, Emmanuel Ifeanyi O, Martin OI, Patience Ebele N, Ogochukwu MTBO. Evaluation of some immunological and haematological indices of hepatitis B infected subjects in Nnamdi Azikiwe University Teaching Hospital, Nnewi, Anambra State, Nigeria. *J Biomed Sci*. 2017;06:1–7.  
DOI: 10.4172/2254-609X.100061
  34. Benninger M, Segreti J. Is it bacterial or viral? Criteria for distinguishing bacterial and viral infections. *J Fam Pract*. 2008;57:S5-11.  
Available: <http://www.ncbi.nlm.nih.gov/pubmed/18662527>
  35. Dou J, Lou Y, Wu J, Lu Y, Jin Y. Thrombocytopenia in patients with hepatitis B virus-related chronic hepatitis: Evaluation of the immature platelet fraction. *Platelets*. 2014;25:399–404.  
DOI: 10.3109/09537104.2013.832742
  36. Nwokediuko SC, Ibegbulam O. Quantitative platelet abnormalities in patients with hepatitis B virus-related liver disease. *Gastroenterol Res. Elmer Press*. 2009;2:344–349.  
DOI: 10.4021/gr2009.12.1329
  37. Hayashi H, Beppu T, Shirabe K, Maehara Y, Baba H. Management of thrombocytopenia due to liver cirrhosis: A review. *World J Gastroenterol*. 2014;20:2595–2605.  
DOI: 10.3748/wjg.v20.i10.2595
  38. Ifeanyi OE, FCO, HAN, KCO, Ogochukwu MTBO, Francisca OU, et al. Impact of HIV and hepatitis B virus coinfection on selected haematological markers of the patients in Umuahia, Abia State, Nigeria. *Ann Clin Lab Res*. 2017;05:9–12.  
DOI: 10.21767/2386-5180.1000175
  39. Mao W, Sun Q, Fan J, Lin S, Ye B. AST to platelet ratio index predicts mortality in hospitalized patients with hepatitis B-related decompensated cirrhosis. *Medicine (Baltimore)*. Wolters Kluwer Health. 2016;95:e2946.  
DOI: 10.1097/MD.0000000000002946
  40. Kim WR, Berg T, Asselah T, Flisiak R, Fung S, Gordon SC, et al. Evaluation of APRI and FIB-4 scoring systems for non-invasive assessment of hepatic fibrosis in chronic hepatitis B patients. *J Hepatol*. 2016;64:773–780.  
DOI: 10.1016/j.jhep.2015.11.012
  41. Murali AR, Carey WD. Liver test interpretation - approach to the patient with liver disease: A guide to commonly used liver tests. In: *Cleveland clinic disease management [Internet]*; 2017. [Cited 2 Dec 2017]  
DOI: 10.1136/bmj.38050.593634.63
  42. Botros M, Sikaris KA. The de Ritis ratio: The test of time. *Clin Biochem Rev. The Australian Association of Clinical Biochemists*. 2013;34:117–30.  
Available: <http://www.ncbi.nlm.nih.gov/pubmed/24353357>
  43. Prati D, Taioli E, Zanella A, Della Torre E, Butelli S, Del Vecchio E, et al. Updated definitions of healthy ranges for serum alanine aminotransferase levels. *Ann Intern Med*. 2002;137:1–10.  
DOI: 10.1093/ajcp/200207020-00006 [pii]
  44. Eminler AT, Ayyildiz T, Irak K, Kiyici M, Gurel S, Dolar E, et al. AST/ALT ratio is

- not useful in predicting the degree of fibrosis in chronic viral hepatitis patients. *Eur J Gastroenterol Hepatol.* 2015;27:1361–1366.  
DOI: 10.1097/MEG.0000000000000468
45. Teshale E, Lu M, Rupp LB, Holmberg SD, Moorman AC, Spradling P, et al. APRI and FIB-4 are good predictors of the stage of liver fibrosis in chronic hepatitis B: The chronic hepatitis cohort study (CHeCS). *J Viral Hepat.* 2014;21:917–920.  
DOI: 10.1111/jvh.12279
46. Nyblom H, Berggren U, Balldin J, Olsson R. High AST/ALT ratio may indicate advanced alcoholic liver disease rather than heavy drinking. *Alcohol Alcohol.* Oxford University Press. 2004;39:336–339.  
DOI: 10.1093/alcalc/agh074
47. Parikh P, Ryan JD, Tsochatzis EA. Fibrosis assessment in patients with chronic hepatitis B virus (HBV) infection. *Ann Transl Med.* AME Publications. 2017;5:40.  
DOI: 10.21037/atm.2017.01.28
48. Li Q, Ren X, Lu C, Li W, Huang Y, Chen L. Evaluation of APRI and FIB-4 for noninvasive assessment of significant fibrosis and cirrhosis in HBeAg-negative CHB patients with ALT $\leq$ 2 ULN: A retrospective cohort study. *Medicine (Baltimore).* Wolters Kluwer Health. 2017;96:e6336.  
DOI: 10.1097/MD.00000000000006336
49. Ma J, Jiang Y, Gong G. Evaluation of seven noninvasive models in staging liver fibrosis in patients with chronic hepatitis B virus infection. *Eur J Gastroenterol Hepatol.* 2013;25:428–434.  
DOI: 10.1097/MEG.0b013e32835cb5dd
50. Béguelin C, Fall F, Seydi M, Wandeler G. The current situation and challenges of screening for and treating hepatitis B in sub-Saharan Africa. *Expert Rev Gastroenterol Hepatol.* 2018;12:537–546.  
DOI: 10.1080/17474124.2018.1474097
51. Fiacre Bagnaka Eloumou SA. Clinicopathological characteristics of a group of Sub-Saharan African patients with chronic hepatitis B infection: A Gender analysis. *Gastroenterol Hepatol Open Access.* 2016;5.  
DOI: 10.15406/ghoa.2016.05.00174

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